

CO₂, N₂O and NH₃ emissions from two different type of soils as affected by applications of dairy sewage sludge**

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A b s t r a c t. A pot experiment with the dairy sewage sludge (DSS) was conducted under aerobic condition (60% WHC, water holding capacity) for a period of 240 days. The emissions of carbon dioxide (CO₂), nitrous oxide (N₂O) and ammonia (NH₃) were determined in grey-brown podzolic and brown soils. Both soils were amended by different doses of DSS (0, 30, 60, 80, 120, 200, 300 and 600 t ha⁻¹). In general, the amendment of DSS stimulated CO₂ and N₂O emissions from both soils. This effect increased after the incorporation of high doses of DSS. It was confirmed by significant positive correlations between the doses of DSS and CO₂ and N₂O fluxes. This study showed that DSS application had no significant influence on the emission of NH₃. In both soils the NH₃ emission was noted only once during the incubation period and only in the treatments with the highest doses of DSS. The long-term addition of DSS on this parameters needs further examination.

K e y w o r d s: brown soil, carbon dioxide, dairy sewage sludge, grey-brown podzolic soil, nitrous oxide

INTRODUCTION

Soils are regarded as the important sources for the production of the carbon dioxide (CO₂) and nitrous oxide (N₂O). These two gases are important greenhouse gases in the atmosphere contributing about 50 and 5% to the global warming, respectively (Lou *et al.*, 2007). Microbial and chemical processes that occur in the soil affect the concentrations of greenhouse gases in the atmosphere (Mosier, 1998). Soil processes contribute about 1% of CO₂ (Chu *et al.*, 2007), 20% of NH₃ and 70% of N₂O (Mosier, 1998). Therefore, there is no doubt that agriculture plays an important role in the global budget of greenhouse gases (Chu *et al.*, 2007). Increasing concentrations of CO₂ in the atmosphere account for greater plant CO₂ assimilation and biomass pro-

duction in many ecosystems (Mosier, 1998). There are two main groups of microorganisms in soil: heterotrophic and autotrophic organisms, that are involved in the CO₂ production in the soil. Most CO₂ produced by heterotrophic soil organisms is respired by microorganisms, such as bacteria, non-mycorrhizal and mycorrhizal fungi and actinomycetes (Kuzyakov, 2006). CO₂ in soils is produced also through various processes, such as biological oxidation of soil organic matter and decomposition of crop residues and wastes incorporated into the soil, such as sewage sludge (Kuzyakov, 2006; Mosier, 1998). Nitrification and denitrification, microbial processes which rely on mineral nitrogen (ammonia and nitrate) as their substrate, are responsible for most of the N₂O produced in soil (Azam *et al.*, 2002; Firestone and Davidson, 1989). The high antropogenic N₂O emission is also result of increased nitrogen input into agricultural soils (Chu *et al.*, 2004).

Land use of the dairy sewage sludge can also affect on the fluxes of nitrogen oxide (N₂O) and carbon dioxide (CO₂). Dairy sewage sludge, which contains high concentrations of plant nutrients, in particular N and P, and the large of organic matter content, is valuable for maintaining or improving soil biological characteristics (Jezierska-Tys and Frąc, 2005; 2006). It is well known that soil applications of organic and nitrogen materials stimulate surface emissions of CO₂ (Kuzyakov, 2006; Masunaga *et al.*, 2007) and N₂O productions (Ambus *et al.*, 2001). It is little known about the fluxes of CO₂, NH₃ and N₂O under the influence of the dairy sewage sludge (DSS). The aim of this study was to evaluate the emissions of these gases (CO₂, NH₃, N₂O) from two different type of soils in response to various doses of dairy sewage sludge applications in pot experiment.

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MATERIAL AND METHODS

The experiment was carried out on two different type of soil: grey-brown podzolic soil and brown soil. The grey-brown podzolic soil used in this study was collected from Piaseczno while brown soil was collected from Krasnystaw, both located at south-east of Poland. The grey-brown podzolic soil had the following granulometric composition: sand fraction 65%, silt fraction 19% and fine slit and clay fraction 16%. The brown soil was formed from silt-loam with 8% sand fraction, 47% slit fraction and 45% clay and slit fraction. Other characteristics of both soils are given in the Table 1. The dairy sewage sludge (DSS) used in the experiment came from the Regional Dairy Cooperative in Krasnystaw. Some chemical properties of this sludge are presented in Table 1.

Table 1. Characteristics of soils and dairy sewage sludge used in the experiment

Parameters	Brown soil	Grey-brown podzolic soil	Dairy sewage sludge (DSS)
pH _{KCl}	6.4	4.8	8.5
C (g kg ⁻¹)	13.5	4.5	400.0
N (g kg ⁻¹)	1.6	0.4	33.2
C/N	8.3	12.5	12.0
Macronutrients (total g kg ⁻¹)			
P	18.3	5.3	11.5
K	26.8	8.7	2.5
Heavy metals (mg kg ⁻¹)			
Zn	28.7	11.7	95.0
Cd	0.16	< 0.16	2.6
Cu	7.16	2.21	19.8
Pb	10.3	7.08	9.0
Ni	10.1	2.98	12.6
Cr	18.4	8.10	36.8
Hg	0.09	0.03	0.22

The soils were collected up to a 20 cm depth from a few different locations to obtain representative samples. Air-dried and sieved (<2 mm) soil (4 kg) was placed in pots and mixed up with different doses of dairy sewage sludge. There were 16 of treatments (8 in the grey-brown podzolic soil and 8 in the brown soil). There were incorporated the same doses of DSS into each soil. Each treatment consisted at three replicates. The dairy sewage sludge were applicated into soils at the following doses: control – no sludge (DSS-0); 30 (DSS-30); 60 (DSS-60); 80 (DSS-80); 120 (DSS-120); 200 (DSS-200); 300 (DSS-300) and 600 t ha⁻¹ (DSS-600). The soil samples were incubated at the 20°C through 240 days.

Each treatment was watered up to 60% of the total water capacity. The analysis were measured after 14, 30, 60, 90, 120 and 240 days after the incorporation of DSS. 5 g of soil was put into the jar (60 cm³). The soil was watered (5 ml) and closed by stopper. The gas samples were evacuated through a septum in the jar lid.

The following methods were used to determine the CO₂ and N₂O emission: according to Włodarczyk (2000) and Horn *et al.* (1994), respectively. The gas samples were analysed for CO₂ and N₂O by a gas chromatograph (Shimadzu GC-14) equipped with TCD and ECD, respectively. These analyses were done in Institute of Agrophysics PAS in Lublin. The emission of NH₃ from soils was measured by the spectrophotometric method of Kim (1973).

Statistical analysis were performed using Statistica 7.1 software. An ANOVA at a probability level of 5% was used to test the significance of the treatment effects. The correlation test was used to estimate the relationships between the examined characteristics.

RESULTS

In comparison with the control, the amendment of DSS obviously stimulated CO₂ emission from the both soils. Generally, CO₂ fluxes from the grey-brown soil were higher than from the brown soil. CO₂ fluxes were affected by the DSS amended to the soils, especially at the early stage of the incubation (Fig. 1). From grey brown podzolic soil the highest CO₂ emission was observed since the beginning of the incubation till 60 days after the incorporation of DSS and then decrease until the end of this study. The little increase was noted only since the last terms of analysis. In brown soil the highest stimulation of CO₂ emission was observed also at the beginning of the incubation till 30 days of the experiment. The stimulating effect showed up better in gas samples from grey-brown podzolic soil than brown soil. The highest increase CO₂ emission from both soils was measured in the treatments with the highest doses of DSS, that is 200, 300 and 600 t ha⁻¹ (Fig. 2). Statistical analyses further indicated that CO₂ fluxes were significantly positively correlated with doses of the DSS incorporated to the soils and N₂O emission from soils. The significantly negative correlation was noted between CO₂ emission and the soils time of incubation (Table 2).

The variations of N₂O emission from soils are presented in Fig. 3. The N₂O emission was varied according to the type of soils. In grey-brown podzolic soil, compared with the control, N₂O emission was rapidly increased from 0 to 30 days after the amendment of DSS and then gradually reduced to the lower values near to the control. In brown soil the stimulation of N₂O fluxes were observed only after 30 days of incubation. In other terms of analysis the N₂O emission was at the same level in all the treatments. Statistical analysis confirmed that only the highest doses of DSS significantly increased the N₂O emission from soils

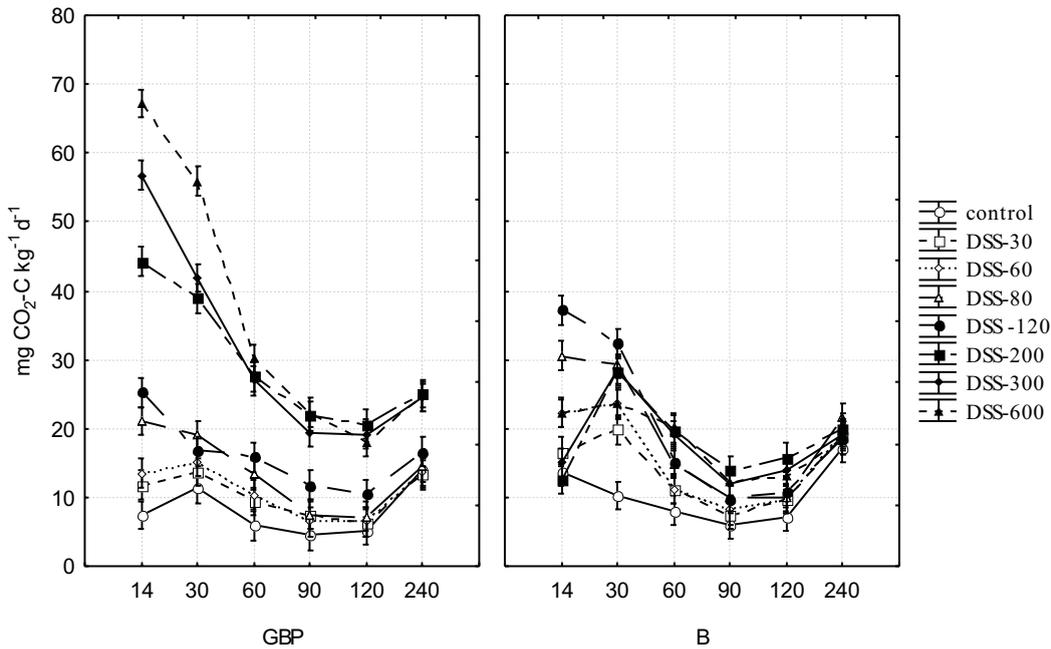


Fig. 1. Changes of CO₂ fluxes from a grey-brown podzolic (GBP) and brown (B) soils amended with dairy sewage sludge under aerobic incubation. Values are the mean of triplicates. Vertical bars indicate the standard error of the averages. ANOVA for dose of DSS – LSD_{0.05}=3.13.

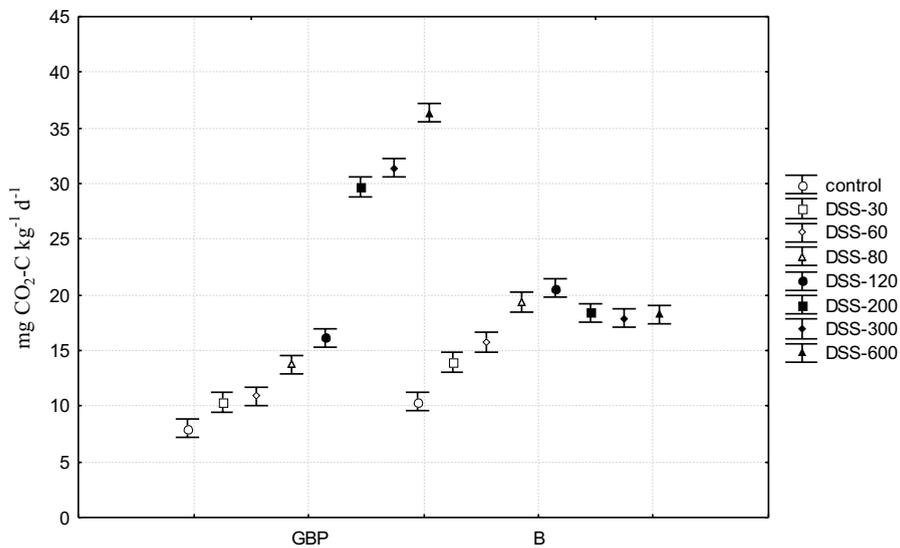


Fig. 2. The mean values of CO₂ fluxes during incubation period.

(Fig. 4). The statistical analyses showed significant positive correlations between the N₂O emission and the dose of DSS and CO₂ fluxes. The negative correlation was observed between N₂O emission and soils incubation time (Table 2).

The content of NH₃ emission from both soils was at the level of zero during incubation time in most of the treatments. In grey-brown podzolic soil, the significant increase of NH₃ emission was observed only 30 days after the in-

corporation of DSS in the treatment with the highest dose of sludge (600 t ha⁻¹). In brown soil, the increase of NH₃ emission was observed also 30 days after the application of DSS in treatments with 300 and 600 t ha⁻¹ (Fig. 5). These values had the influence on the level of means values of NH₃ emission, that are presented on the Fig. 6. Amounts of NH₃ emitted were significantly positively correlated with the dose of DSS and negatively with incubation time (Table 2).

Table 2. Statistical characteristic (correlation coefficients)

Parameters	DSS dose	Incubation time	CO ₂	N ₂ O	NH ₃
DSS dose	–	–	0.54 ***	0.33 ***	0.22 ***
Incubation time		–	-0.36 ***	-0.33 ***	-0.20 ***
CO ₂			–	0.72 ***	0.14 *
N ₂ O				–	n.s.
NH ₃					–

DSS – dose of dairy sewage sludge. Indicate significance at the: *5, **1, ***0.1% level, n.s. – no significant.

emission from soil amended with sludge suggests that soil microbial activity is affected by this waste application in a prolonged period. Similar results were obtained in the presented study, but the dairy sewage sludge was used in this experiment.

Nitrous oxide fluxes are often stimulated by mineral fertiliser applications, particularly on wet soil (Clayton *et al.*, 1994), and also by organic amendment (Ball *et al.*, 2002). In our research DSS applications increased the emission of N₂O. However, the incorporation of DSS had little effect on the N₂O emission, because the single application resulted in increase only at the beginning of the experiment. The results of the present study do not indicate that losses of N₂O are high during single application of DSS, but N₂O emission increases with the dose of this waste application.

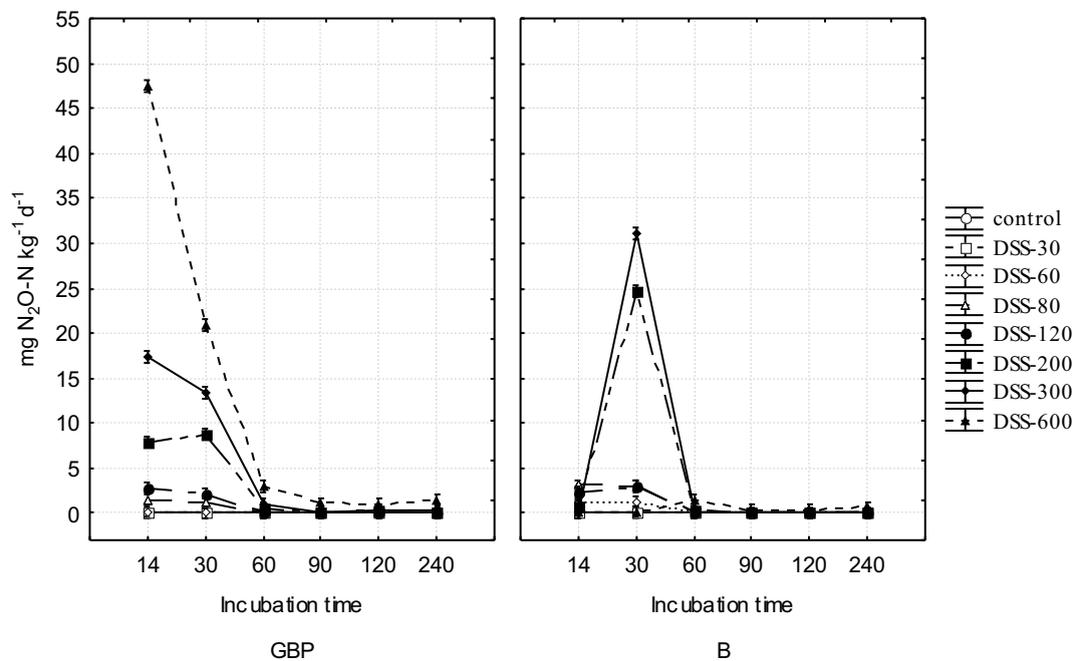


Fig. 3. Changes of N₂O fluxes from a grey-brown podzolic (GBP) and brown (B) soils amended with dairy sewage sludge under aerobic incubation. ANOVA for dose of DSS – LSD_{0.05}=9.93.

DISCUSSION

Under non-planted conditions, soil CO₂ emission is mainly caused by microbial respiration, and it is important parameter for evaluating microbial activity in soils (Lou *et al.*, 2007). In our study CO₂ emission was significantly higher in the dairy sewage sludge amended soils than in the control (Fig. 1). Soil CO₂ flux was rapidly increased at the beginning of the experiment. This indicated that the microbial activities were stimulated by probably the increased C supply and the substrates after DSS incorporation. Significantly positive correlation was obtained between CO₂ flux and dose of DSS, with the correlation coefficient 0.54***. According to Ambus *et al.* (2001) the high content of CO₂

According to Ambus *et al.* (2001) the utilisation of sewage sludge in agriculture may have a relatively limited impact on the N₂O gas balance. Nitrous oxide emissions measured in the soil with different doses of DSS indicated that organic matter and nitrogen incorporated into soil with this sludge stimulated microbial activity, especially in the treatments with high doses of the DSS. A significant positive correlations between N₂O and CO₂ emissions observed in the present study and these reported by others (Azam *et al.*, 2002) support the contention that enhanced microbial activity is responsible, at least partially, for the observed N₂O emissions that followed nitrogen matter addition.

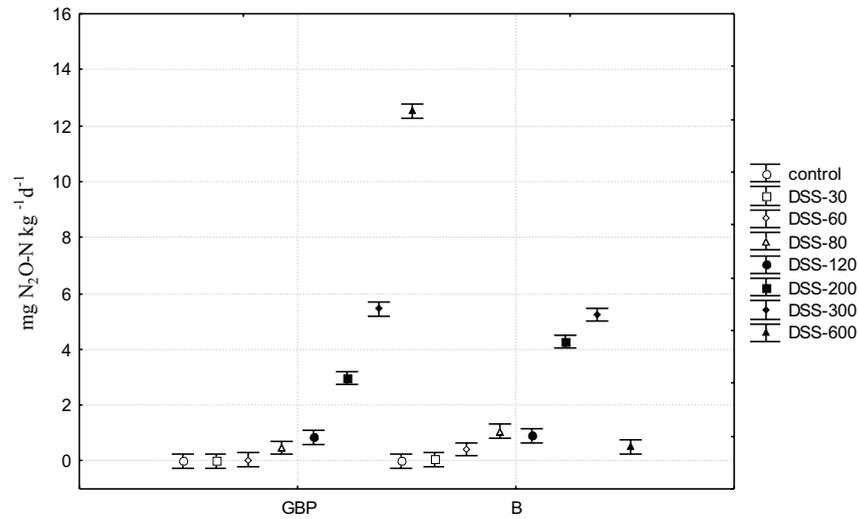


Fig. 4. The mean values of N₂O fluxes during incubation period.

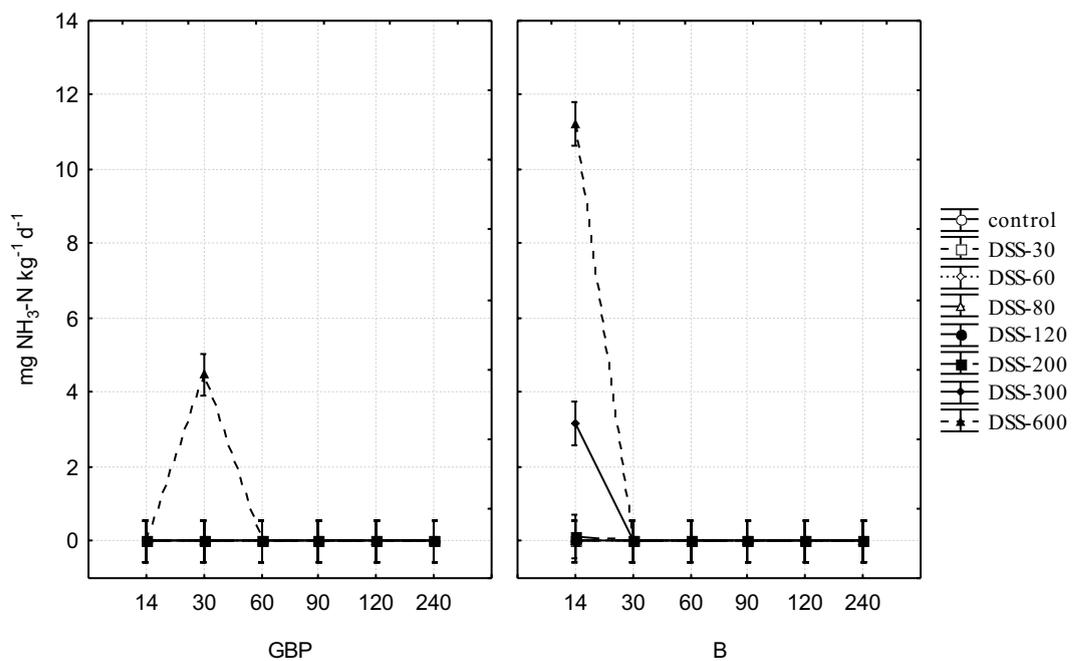


Fig. 5. Changes of NH₃ emission from a grey-brown podzolic (GBP) and brown (B) soils amended with dairy sewage sludge under aerobic incubation. ANOVA for dose of DSS – LSD_{0.05}=0.85.

In our experiment only one type of organic wastes (DSS) was tested, further research is needed to investigate the contribution of different wastes to the emissions of CO₂ and N₂O, and to assess the controlling factors to predict CO₂ and N₂O emissions from different type of soils. It is important to assess how the long-term addition of DSS will influence on the CO₂ and N₂O emissions, because in this study only single application was researched. This needs further examination.

CONCLUSIONS

1. The present study showed that DSS application, in general, had no significant effect on the emission of NH₃ from soils. In both soils the NH₃ emission was noted only once during the incubation period in the treatments with the highest doses of DSS (300 and 600 t ha⁻¹).
2. The results of our research indicated that the dairy sewage sludge is one kind of the organic wastes that increased emission of greenhouse gases (CO₂, N₂O) from soils.

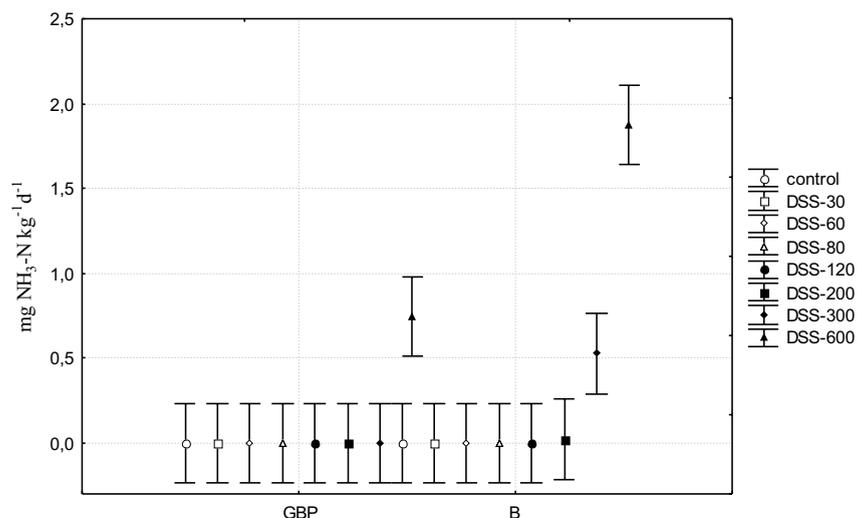


Fig. 6. The mean values of NH₃ emission during incubation period.

REFERENCES

- Ambus P., Jensen J.M., Prieme A., Pilegaard K., and Kjoller A., 2001. Assessment of CH₄ and N₂O fluxes in a Danish beech (*Fagus sylvatica*) forest and an adjacent N-fertilised barley (*Hordeum vulgare*) field: effects of sewage sludge amendments. *Nutr. Cycl. Agroecosys.*, 60, 15-21.
- Azam F., Müller C., Weiske A., Benckiser G., and Ottow J.C.G., 2002. Nitrification and denitrification as sources of atmospheric nitrous oxide – role of oxidizable carbon and applied nitrogen. *Biol. Fert. Soils*, 35, 54-61.
- Ball B.C., McTaggart I.P., and Watson C.A., 2002. Influence of organic ley-arable management and afforestation in sandy loam to clay loam soils on fluxes of N₂O and CH₄ in Scotland. *Agr. Ecosyst. Environ.*, 90, 305-317.
- Chu H., Hosen Y., and Yagi K., 2004. Nitrogen oxide emissions and soil microbial properties as affected by N-fertilizer management in a Japanese Andisol. *Soil Sci. Plant Nutr.*, 50, 287-292.
- Chu H., Hosen Y., and Yagi K., 2007. NO, N₂O, CH₄ and CO₂ fluxes in winter barley field of Japanese Andisol as affected by N fertilizer management. *Soil Biol. Biochem.*, 39, 330-339.
- Clayton H., Arah J.R.M., and Smith K.A., 1994. Measurement of nitrous oxide emissions from fertilised grassland using closed chambers. *J. Geophys. Res.*, 99, 16599-16607.
- Firestone M.K. and Davidson E.A., 1989. Microbiological basis of NO and N₂O production and consumption in soil. In: *Exchange of Trace Gases Between Terrestrial Ecosystems and the Atmosphere* (Eds M.O. Andreae and D.S. Schimel). Wiley, Chichester, England.
- Horn R., Stępniewski W., Włodarczyk T., Walenzik G., and Eckhardt F.E.W., 1994. Denitrification rate and microbial distribution within homogeneous model soil aggregates. Microbial distribution within soil aggregates. *Int. Agrophysics*, 8, 65-74.
- Jezierska-Tys S. and Frąc M., 2005. The effect of fertilization with sewage sludge from a dairy plant and with rape straw on the population numbers of selected microorganisms and respiration activity of brown soil. *Polish J. Soil Sci.*, 38, 145-151.
- Jezierska-Tys S. and Frąc M., 2006. Enzymatic activity of grey-brown podzolic soil enriched with sewage sludge from a dairy plant. *Polish J. Soil Sci.*, 39, 33-42.
- Kim Ch.M., 1973. Influence of vegetation types on the intensity of ammonia and nitrogen dioxide liberation from soil. *Soil Biol. Biochem.*, 5, 163-166.
- Kuzyakov Y., 2006. Sources of CO₂ efflux from soil and review of partitioning methods. *Soil Biol. Biochem.*, 38, 425-448.
- Lou Y., Ren L., Li Z., Zhang T., and Inubushi K., 2007. Effect of rice residues on carbon dioxide and nitrous oxide emissions from a paddy soil of subtropical China. *Water Air Soil Poll.*, 178, 157-168.
- Masunaga T., Sato K., Senga Y., Seike Y., Inaishi H., and Wakatsuki T., 2007. Characteristics of CO₂, CH₄ and N₂O emissions from a multi-soil-layering system during wastewater treatment. *Soil Sci. Plant Nutr.*, 53, 173-180.
- Mosier A.R., 1998. Soil processes and global change. *Biol. Fert. Soils*, 27, 221-229.
- Włodarczyk T., 2000. N₂O emissions and absorption against a background of CO₂ in eutric cambisol under different oxidation-reduction conditions (in Polish). *Acta Agrophysica*, 28, 1-132.